



Research Article

Relation between Water Filled Pore Space, $\text{NO}_3\text{-N}$ and N_2O Emission from Wheat Cropped soil

AMITA RAJ¹, ARTI BHATIA^{1*}, PRAGYA², BIDISHA CHAKRABARTI¹, NIVETA JAIN¹, V.K. SEHGAL², Y.S. SHIVAY³ AND RITU TOMAR¹

¹Centre for Environmental Science and Climate Resilient Agriculture, ²Division of Agricultural Physics, ³Division of Agronomy, ICAR-Indian Agricultural Research Institute, New Delhi-110012, India

ABSTRACT

Greenhouse gas (GHG) emissions contribute enormously to climate change. N_2O is a crucial greenhouse gas emitted from crop fields and is affected by management and soil properties of the crop field. Agronomical management practices such as irrigation profoundly affects nitrous oxide (N_2O) emissions from soil. Proper irrigation management considering soil type helps in mitigating N_2O emissions from crop field soils. Also, N-fertilizer's use has direct influence on N_2O emission from managed soils. Therefore, in the present study, N_2O emissions from wheat crop (HD2967) were examined in relation to the irrigation provided and nitrogenous fertilizer applied to wheat crop grown in two *rabi* seasons (2015-2017). Positive correlation was observed between water filled pore spaces and N_2O emission from wheat crop field for both the growing seasons (2015-17). Positive correlation was also observed between $\text{NO}_3\text{-N}$ and N_2O emission indicating nitrogenous fertilizer application increases N_2O emission in wheat crop.

Key words: Nitrous oxide, Climate Change, Water filled pore spaces, Wheat

Introduction

Agriculture contributes to climate change by emitting greenhouse gases. However, because they are interconnected, it is also influenced by climate change, both directly and indirectly. Significant volumes of greenhouse gases (GHGs) are discharged into the atmosphere, with soil management and livestock accounting for 14 percent of the total. Nitrous oxide (N_2O) plays role in climate-alteration gases contributing 5%, to escalate global warming (Watson *et al.*, 1996). Nitrous oxide (N_2O) is produced in agricultural soil because of microbial nitrification and denitrification (Bhatia *et al.*, 2012a). The rate of N_2O emission from crop field is greatly influenced by many factors which includes soil

temperature, soil moisture, soil air and carbon availability, type of crop, residue management and application of nitrogenous fertilizers.

Nitrous oxide emissions from wheat crop fields in Trans Indo-Gangetic Plains ranged between 0.31 and 0.98 kg $\text{N}_2\text{O-N}$ /ha/year, depending on fertilizer and irrigation treatments (Gupta *et al.*, 2016). The rice-wheat system of the Indo-Gangetic plains resulted in $\text{N}_2\text{O-N}$ emissions of 1.57 kg/ha, or 0.38 percent of applied N when N applied by urea was 240 kg N/ha/year (Pathak *et al.*, 2002). It has been estimated that N_2O emission from wheat fields in India is 0.17 Tg/year which is a significant cause of climate change (Bhatia *et al.*, 2012b). N_2O emissions are so much influenced by availability of N in soil which in turn is affected by chemical fertilizers present in the agricultural soils (Cowan *et al.*, 2021;

*Corresponding author,
Email: artibhatia.iari@gmail.com



Fig. 1. Pictorial representation of experimental plot with placement of closed chambers for gas sampling at MB-14 B experimental farm of the Centre for Environment Science and Climate Resilient Agriculture (CESCRA) division in IARI

Fagodiya *et al.*, 2019). Another factor influencing N_2O emission is soil moisture due to its ability to regulate availability of oxygen to soil microorganisms (Davidson *et al.*, 2000). Crop management and soil conditions have an impact on greenhouse gas (GHG) emissions from crop fields. Irrigation is one such activity that has a significant impact on soil nitrous oxide (N_2O) emissions (Bhatia *et al.*, 2010). As a result, efficient irrigation management combined with crop field soil attributes might reduce N_2O emissions from agricultural soils.

Materials and Method

A field experiment was carried out during (2015-16 and 2016-17) growing wheat (*Triticum aestivum* L.) in the *rabi* season, at the MB-14 B experimental farm of the Centre for Environment Science and Climate Resilient Agriculture (CESCRA) division in IARI, New Delhi, India ($28^{\circ}35' N$ and $77^{\circ}12' E$), in the trans Indo-Gangetic alluvial tract which is located at an elevation of 228 meters above mean sea level (AMSL) (Fig.1). New Delhi is situated in northern part of India, and it has subtropical and semi-arid type climate. The mean annual maximum air temperature is $35^{\circ}C$ and the mean annual minimum air temperature is $18^{\circ}C$. The mean monthly minimum and maximum temperature ranges from ~ 4 to $6^{\circ}C$ in January and $\sim 40^{\circ}C$ to $45^{\circ}C$ in the month of June, respectively. The area receives ~ 650 to 750 mm annual rainfall, up to 80-85 % occurring from June to September. Soil type is Ustochrept with pH 8.6

and texture is sandy loam (21% clay, 33% silt and 46% sand). Wheat crop (HD 2967) was directly sown during *Rabi* season on 30th November, 2015. The plant to plant spacing was 5cm and row to row spacing was 20 cm with plant density of 25 plants m^{-2} . The crop was irrigated with proper time interval keeping in mind all the critical stages. Nitrogen application rate was 120 kg ha^{-1} in 3 splits doses, first 50% as basal and next 50% in two equal halves (25% each) at 21 DAS and 52 DAS and 87 DAS as top dressing. At the time of sowing, a base dosage of P and K was administered at a rate of 60 kg ha^{-1} . All the inter-cultural practices in both the crops such as sowing, weeding, and thinning management as well as crop pest control activities were taken care of as per standard procedures. Weeding was done manually, and no chemicals (herbicide and pesticide) were used to avoid their added effects. Treatment details of the crop are mentioned in Table 1.

The collection and sampling of GHGs from the experimental site was performed by adopting closed chamber technique (Malyan *et al.*, 2019; Bhatia *et al.*, 2005). The chambers used were made of acrylic sheet of 6 mm thickness with a standard box dimension of $50\text{ cm} \times 30\text{ cm} \times 100\text{ cm}$ (length \times breadth \times height). Chamber (box) was fitted with battery operated fan to homogenize its inner air and temperature monitored by a thermometer was kept inside the chamber. To collect gas samples from the chamber, a three-way stopcock fitted at the top was attached with rubber septum (Eastern Medikit Ltd. India). Aluminum channels were utilized as a base for placing the acrylic chamber points and were randomly placed in the field at the sampling points. After installing the chamber above it, the aluminum channels were pushed into the soil to a depth of 10 cm and then filled with water to create an airtight system. The gas samples were taken from the chambers using hypodermic needle of 24 gauge fitted with a 50 mL syringe. The chambers were flushed numerous times with the syringe before taking the sample. The gas sampling was carried out in the morning time and three samples were drawn from each chamber at 0, 1/2 and 1 hrs. between 9:00 and 11.00 A.M. After collecting each sample, syringes were air tightened with a three-way stopcock and returned to the laboratory within 24 hours for further examination of GHG concentrations. The head space

Table 1. Wheat crop field treatment specifics

Treatment	Wheat crop	
	2015 -2016	2016-2017
Variety	HD 2967	HD 2967
Date of Sowing	30.11.2015	02.12.2016
Irrigation schedule (Days after sowing- DAS)	<ul style="list-style-type: none"> • Average Irrigation amount: - 50-60 mm • No. of irrigation: - 5 <ul style="list-style-type: none"> • 20 DAS • 40 DAS • 60 DAS • 75 DAS • 95 DAS 	
Fertilizer scheduling	NCU	DAS
• NCU (split doses) @120 kg/ha	1. 50% -	21
• P&K @ 60 Kg/ha	2. 25% -	52
	3. 25% -	87
	P&K	
	100% basal dose	

volume inside the chamber was also measured to quantify the site's GHG flow. Similarly, samples were drawn once in a week (7 days regular interval) throughout the cropping season (Bhatia *et al.*, 2012b). The concentrations of N₂O in the collected gas samples were measured in the research laboratory using Gas Chromatographs (GC) (Hewlett Packard 5890 Series II) equipped with different detectors such as electron capture detector (ECD) and flame ionisation detector (FID), and a stainless-steel column (6 1/8- Porapak N). The carrier gas in GC was N₂, which flowed at a rate of 14 ml min⁻¹. The standards of GHGs, used for calibration, were obtained from Spectra Gases (NIST standards), USA.

The concentration of N₂O in the gas samples was determined using a GC coupled with an ECD (GC-ECD). The temperatures of the injector, column, and detector were kept constant at 120°C, 50°C, and 350°C, respectively. For gaseous calibration, the standards (NIST standards) used were 500 ppbv and 1 ppm for N₂O. The peak area was plotted and measured using a GC-computer interface and a Hewlett Packard integrator. The N₂O concentration in a gas sample was calculated using the method described by Pathak *et al.* (2003). The estimation of total N₂O emission during both the crop seasons was carried out by successive linear interpolation of mean

emissions of a particular GHG during the sampling days of the cropping season assuming that emissions followed a linear trend during the non-sampling periods (Malyan *et al.*, 2021a; 2021b).

Nitrate in Soil

For estimating nitrate nitrogen (NO₃-N) in the soil, we followed standard procedure of the Keeney and Nelson (1982).

Water Filled Pore Space (WFPS)

It is the ratio of volumetric soil moisture content to total soil porosity. Since particle and water densities are consistent at 2.6 and 1 kg/l, the WFPS of soil was calculated simply by combining its bulk density and soil moisture content. It was calculated using the formula as given below:

$$\text{WFPS} = [\text{Gravimetric moisture content} \times \text{Soil bulk density}] / \text{Total soil porosity}$$

Where,

$$\text{Soil porosity} = 1 - [\text{soil bulk density} / \text{soil particle density}]$$

Soil particle density was estimated by pycnometer method and the bulk density was estimated using cylinder method (Blake and Hartge, 1986).

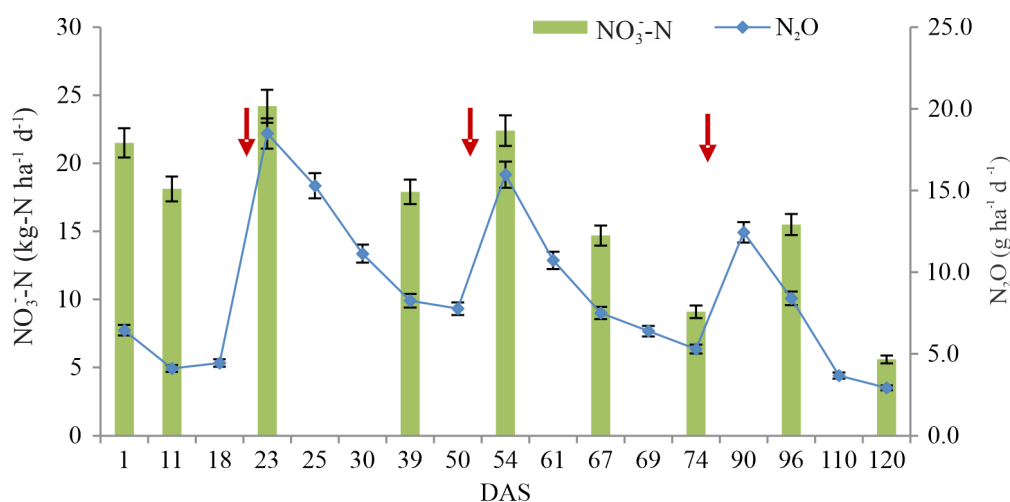


Fig. 2. Relation between N₂O emission and NO₃⁻-N in wheat field for both rabi seasons (2015-17)

Results and Discussion

In our study, the highest NO₃⁻-N was observed during the peak flux of N₂O emission (23 DAS). This coincided with the maximum value of NO₃⁻-N in soil (Fig. 2) which was due to fertilizer application on 20 DAS. This shows that application of nitrogenous fertilizer resulted in the increase in soil ammonium content due to hydrolysis of urea in the presence of high WFPS and with time the partially oxidized condition led to nitrification of the soil ammonium to nitrate rapidly (Banerjee *et al.*, 2002). Thus, higher nitrate content in the soil lead to a comparatively high value of N₂O emission indicating use of N-fertilizers has a direct influence on N₂O emission (Zanatta *et al.*, 2010). Moreover, with a decrease in NO₃⁻-N availability in soil, the N₂O emission also correspondingly declined in a similar trend due to reduced denitrification (Cavuela *et al.*, 2013). The maximum N₂O emission coincided with the maximum WFPS value and NO₃⁻-N in soil. Three peaks of N₂O emission were observed during the crop season, which were 18.5, 16.0 and 12.5 N₂O (g ha⁻¹d⁻¹) at 23, 54 and 90 DAS (cumulative data), respectively. The corresponding WFPS values on these days were 75.8%, 68% and 61%, respectively. Higher value of soil water content leads to increase in rate of N₂O emission mainly due to limited oxygen diffusion in soil through soil pores (Bhattacharya *et al.*, 2018). Baggs and Bateman (2005) reported through their experiment that nitrification rate increases with increase in WFPS but after 60% WFPS

denitrification dominates which is mainly due to reduced availability of substrate (O₂ and CO₂) for nitrifiers caused by restricted rates of diffusion resulting in anaerobic condition. Thus, beyond 60% of WFPS value denitrification process is usually associated with soil moisture content. Positive correlation (R² = 0.55) was found between WFPS and N₂O (Fig. 4) indicating increase in WFPS will lead to anaerobic condition suitable for denitrification which can be supported by similar research report of Garcia-Marco *et al.* (2014) which states that the percentage of WFPS was the primary factor associated in N₂O emission and must be kept below 80% to reduce emissions. Similarly, positive correlation (R² = 0.63) was observed between NO₃⁻-N and N₂O (Fig. 3) which indicates direct effect of nitrogenous fertilizer application on N₂O emission.

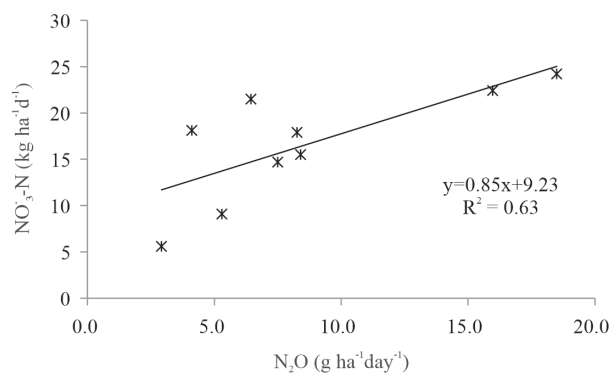


Fig. 3. Relation between N₂O emission and NO₃⁻-N in wheat field for both rabi seasons (2015-17)

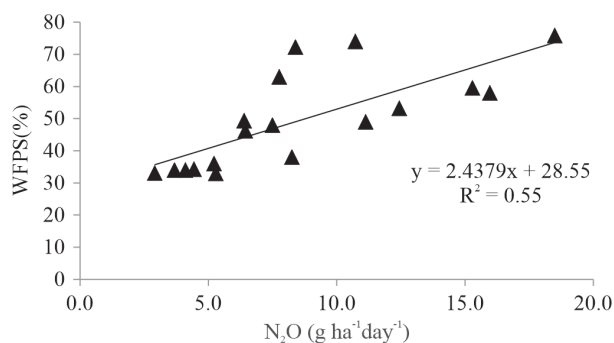


Fig. 4. Relationship between WFPS and N₂O emission from wheat crop for both *rabi* seasons (2015-17)

Conclusion

Through our study, it was observed that both nitrogenous fertilizer application and irrigation scheduling facilitates N₂O production and emission from wheat crop. The nitrogen fertilizer should be applied as per crop requirement, to enable better N uptake by plant and reduce the NO₃ availability for loss as N₂O through denitrification. The irrigation scheduling should not coincide with a fertilizer N application event as anaerobic conditions will increase the WFPS and NO₃ will undergo denitrification. Therefore, judicious scheduling of nitrogenous fertilizer application along with time of irrigation management will avoid the favorable condition for N₂O emission from soil and could be one of the mitigating measures for N₂O emission from fertilized crop soils.

Acknowledgement

The authors express gratitude to the PG School and Director, ICAR-Indian Agricultural Research Institute, New Delhi, for providing the necessary facilities and financial support, for conducting this research work. Financial assistance provided by the NICRA project is also acknowledged.

References

Bateman, E.J. and Baggs, E.M. 2005. Contributions of nitrification and denitrification to N₂O emissions from soils at different water-filled pore space. *Biology and Fertility of Soils* **41**(6): 379-388.

Bhatia, A., Pathak, H., Jain, N., Singh, P.K. and Singh, A.K. 2005. Global warming potential of manure amended soils under rice-wheat system in the

Indo-Gangetic plains. *Atmospheric Environment* **39**(37): 6976-6984.

- Bhatia, A., Aggarwal, P.K., Jain, N. and Pathak, H. 2012b. Greenhouse gas emission from rice and wheat growing areas in India: spatial analysis and upscaling. *Greenhouse Gases: Science and Technology* **2**(2): 115-125.
- Bhatia, A., Pathak, H., Jain, N., Singh, P.K. and Tomer, R. 2012a. Greenhouse gas mitigation in rice-wheat system with leaf color chart-based urea application. *Environmental Monitoring and Assessment* **184**(5): 3095-3107.
- Bhatia, A., Pathak, H., Aggarwal, P.K. and Jain N. 2010. Trade-off between productivity enhancement and global warming potential of rice and wheat in India. *Nutr. Cycl. Agroecosyst.* **86**: 413-424. <https://doi.org/10.1007/s10705-009-9304-5>
- Bhattacharyya, R., Bhatia, A., Das, T.K., Lata, S., Kumar, A., Tomer, R. and Biswas, A.K. 2018. Aggregate-associated N and global warming potential of conservation agriculture-based cropping of maize-wheat system in the north-western Indo-Gangetic Plains. *Soil and Tillage Research* **182**: 66-77.
- Blake, G.R. and Hartge, K.H. 1986. Bulk Density Methods of Soil Analysis, Part 1, Physical and Mineral Method. Ed. by A. Klute, 363-382.
- Bouwman, A.F., Boumans, L.J.M. and Batjes, N.H. 2002. Emissions of N₂O and NO from fertilized fields: Summary of available measurement data. *Global Biogeochemical Cycles* **16**(4): 6-1.
- Cayuela, M.L., Sánchez-Monedero, M.A., Roig, A., Hanley, K., Enders, A. and Lehmann, J. 2013. Biochar and denitrification in soils: when, how much and why does biochar reduce N₂O emissions?. *Scientific Reports* **3**(1): 1-7.
- Davidson, E.A., Keller, M., Erickson, H.E., Verchot, L.V. and Veldkamp, E. 2000. Testing a conceptual model of soil emissions of nitrous and nitric oxides: using two functions based on soil nitrogen availability and soil water content, the hole-in-the-pipe model characterizes a large fraction of the observed variation of nitric oxide and nitrous oxide emissions from soils. *Bioscience* **50**(8): 667-680.
- Fagodiya, R.K., Pathak, H., Bhatia, A., Jain, N., Gupta, D.K., Kumar, A. and Tomer, R. 2019. Nitrous oxide emission and mitigation from maize-wheat

- rotation in the upper Indo-Gangetic Plains. *Carbon Management* **10**(5): 489-499.
- García Marco, S., Ravella, S.R., Chadwick, D., Vallejo, A., Gregory, A.S. and Cárdenas, L.M. 2014. Ranking factors affecting emissions of GHG from incubated agricultural soils. *European Journal of Soil Science* **65**(4): 573-583.
- Gupta, D., Bhatia, A., Das, T.K., Singh, P., Kumar, A., Jain, N. and Pathak, H. 2016. Economic analysis of different greenhouse gas mitigation technologies in rice-wheat cropping system of the Indo-Gangetic Plains. *Current Science* **110**: 867-874. <http://www.jstor.org/stable/24907971>
- Pathak, H., Bhatia, A., Prasad, S., Singh, S., Kumar, S., Jain, M.C. and Kumar, U. 2002. Emission of nitrous oxide from rice-wheat systems of Indo-Gangetic plains of India. *Environmental Monitoring and Assessment* **77**(2): 163-178.
- Pathak, H., Prasad, S., Bhatia, A., Singh, S., Kumar, S., Singh, J. and Jain, M.C. 2003. Methane emission from rice-wheat cropping system in the Indo-Gangetic plain in relation to irrigation, farmyard manure and dicyandiamide application. *Agriculture, Ecosystems and Environment* **97**(1-3): 309-316.
- Mosier, A.R. and Hutchinson, G.L. 1981. *Nitrous oxide emissions from cropped fields* (Vol. 10, No. 2, pp. 169-173). American Society of Agronomy, Crop Science Society of America, and Soil Science Society of America.
- Keeney, D.R. and Nelson, D.W. 1982. Inorganic forms of nitrogen. In A.L. Page (ed.), *Methods of soil analysis, Part 2*. Edn 2, Vol. IX, Agronomy, 643-698.
- Malyan, S.K., Bhatia, A., Kumar, S.S., Fagodiya, R.K., Pugazhendhi, A. and Duc, P.A. 2019. Mitigation of greenhouse gas intensity by supplementing with Azolla and moderating the dose of nitrogen fertilizer. *Biocatalysis and Agricultural Biotechnology*. <https://doi.org/10.1016/j.bcab.2019.101266>
- Malyan, S.K., Bhatia, A., Fagodiya, R.K., Kumar, S.S., Kumar, A., Gupta, D.K. and Pathak, H. 2021. Plummeting global warming potential by chemicals interventions in irrigated rice: A lab to field assessment. *Agriculture, Ecosystems & Environment* **319**: 107545.
- Malyan, S.K., Bhatia, A., Tomer, R., Harit, R C., Jain, N., Bhowmik, A. and Kaushik, R. 2021. Mitigation of yield-scaled greenhouse gas emissions from irrigated rice through Azolla, Blue-green algae, and plant growth-promoting bacteria. *Environmental Science and Pollution Research* 1-15.
- Shakoor, A., Xu, Y., Wang, Q., Chen, N., He, F., Zuo, H. and Yang, S. 2018. Effects of fertilizer application schemes and soil environmental factors on nitrous oxide emission fluxes in a rice-wheat cropping system, east China. *PLoS one*, **13**(8): e0202016.
- Watson, R.T., Zinyowera, M.C. and Moss, R.H. 1996. *Climate change 1995. Impacts, adaptations and mitigation of climate change: scientific-technical analyses*.
- Wrage, N., Velthof, G.L., Van Beusichem, M.L. and Oenema, O. 2001. Role of nitrifier denitrification in the production of nitrous oxide. *Soil Biology and Biochemistry* **33**(12-13), 1723-1732.
- Zanatta, J.A., Bayer, C., Vieira, F.C., Gomes, J. and Tomazi, M. 2010. Nitrous oxide and methane fluxes in South Brazilian Gleysol as affected by nitrogen fertilizers. *Revista Brasileira de Ciência do Solo* **34**: 1653.

Received: January 14, 2022; Accepted: March 30, 2022